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Obatala Sciences' Protocol 305 How Do I Create ObaGel[®] Cultures for *In Vivo* Implantation?

Written by: Obatala Sciences' Scientific Team Last Updated: January 2024

Reagents, Materials, and Equipment

- Obatala Sciences' ObaGel[®] (Catalog #OS-301)
- ◆ Obatala Sciences' ObaVate[™] (Catalog #OS-302)
- Obatala Sciences' Human Adipose-Derived Stromal/Stem Cells (Catalog #OS-101), Human Stromal Vascular Fraction Cells (Catalog # OS-107-01), or equivalent cryopreserved primary cell product
- Obatala Sciences' StromaQual[™] Stromal Medium (Catalog #OS-001) or medium of choice
- ◆ AdipoQual3D[™] Adipogenic Differentiation Medium (Catalog #OS-310)
- Viability stains such as trypan blue or acridine orange/propidium iodide
- 70% ethanol
- 37°C, 5% CO₂ incubator
- BSL2 Biological Safety Cabinet
- Sterile paper towel or kimwipe
- Conical centrifuge tube
- Micropipette and tips
- Well plate and micropipette for desired format, protocol is optimized for 24-well or 96-well plate
- Wet ice for prolonged handling
- Syringe and needle
- Styrofoam or insulated container

Protocol

Initial Handling of Your Obatala Sciences' Products

- When you receive the package containing your ObaGel[®] and ObaVate[™] products, they may arrive on wet ice or dry ice depending on the shipping conditions.
 - a. Prior to use, thaw the unopened products overnight at 4°C until completely thawed. Do not thaw at room temperature or attempt to warm products at higher temperatures.
 - b. Aliquot necessary volumes for immediate use into separate containers to avoid repeated freeze/thaw cycles.

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- c. After thawing, ObaGel[®] can be stored at 4°C for <48 hours prior to use. For longer term storage up to 3 months, store at -20°C. For storage >3 months, store at -80°C (shelf life 1 year).
- d. After thawing, ObaVate[™] can be stored at 4°C for up to 4 weeks, <3 months at -20°C, and up to 2 years at -80°C.
- 2. After thawing, you may notice protein precipitant present in ObaGel[®]. This is normal and does not impact function or quality of the product. In fact, it is beneficial to the formation of 3D constructs!

Note: Do not attempt to spin or otherwise remove the precipitant from the product, as it will clump and aggregate. Unaltered, the precipitant will disperse when pipetting.

Seeding Cells in ObaGel® for 3-Dimensional Cultures

- 1. Thaw ObaGel[®] and ObaVate[™] overnight at 4°C until they are completely thawed. Do not warm to 37°C. Keep reagents on wet ice while in use.
- 2. Warm growth medium of choice to 37°C.
- 3. Prepare a 50 mL conical tube with 4 mL of growth medium per vial of cells.
- 4. Thaw the cryovial of cells until the moment the ice crystals disappear from the vial.
- 5. Transfer the cryovial to a BSL2 BSC and add the contents of the cryovial dropwise to the 4 mL of growth medium in the conical tube.
- 6. Homogenize the cell suspension by pipetting several times.
- 7. Perform a cell count according to your laboratory's standard operating procedures.
- 8. Centrifuge the conical tube at 1200 RPM for 5 minutes at room temperature.
- Aspirate the supernatant from the cell pellet and resuspend the cells in ObaVate™ at 1 million cells per mL.
- 10. From the 1 million cells per mL solution, calculate the total number of cells needed to create the seeding solution.
- 11. Calculate the volume of concentrated cell solution corresponding to the total cell number from above.
- 12. Subtract the volume of concentrated cell solution from the total volume of ObaVate[™] needed for the seeding solution.¹
- 13. Transfer the volume of concentrated cell solution to a new 50 mL conical tube.
- 14. Add the remaining volume of ObaVate[™] needed for the seeding solution.
- 15. Pipette the cell solution multiple times to mix well.
- 16. Using a micropipette, vigorously pipette the ObaGel® prior to use.
- 17. Add the total volume of ObaGel[®] needed for the seeding solution to the 50 mL conical tube containing the ObaVate[™] cell solution such that the seeding solution contains one-part ObaGel[®] to three-parts ObaVate[™] (1:3 ratio). Pipette several times with a p1000 pipette to mix well.

¹ Obatala recommends accounting for pipetting error by increasing the volume by 10%.

- 18. Once the components are added, quickly transfer the necessary seeding volume of cell suspension onto the tissue culture plate growth surface. Pipette the solution between replicates to maintain homogeneous suspension.^{2 3}
- 19. After aliquoting, transfer the plate to a humidified 5% CO₂ incubator at 37°C. Observe after 1 hour for initial signs of gelation, and again after 24 hours to visualize cell growth.

Implantation of 3-Dimensional ObaGel® constructs

- 1. When transporting ObaGel[®] constructs to the vivarium, do not place on ice. Wrap plates in aluminum foil or appropriately insulated material to aid in temperature maintenance. Place inside a Styrofoam container.
- 2. After anesthetizing the mice, prepare for injections.
- 3. Aspirate ObaGel[®] constructs from the individual wells into syringes by removing the needle from the syringe first, aspirating the ObaGel[®] construct, and finally twisting the needle back onto the syringe for injection.
- 4. Immediately inject the ObaGel[®] constructs into the dorsal subcutaneous top, bottom, left, and right locations (4 locations, 4 injections per mouse).

Recommended Protocols

Obatala Sciences' Protocol 101-How Do I Thaw Cryovials of Cells from Obatala Sciences?

Obatala Sciences' Protocol 301–How Do I Create 3D Cultures with ObaGel®?

We expect that you will have new ideas on how to use our product that extend beyond these boundaries and look forward to hearing about novel ways you can use ObaGel[®] in your discovery research. Please share your findings with us when they become available.

Remember, any laboratory that mentions Obatala Sciences' products by name in a publication is eligible for a 10% discount on their next order! We appreciate not only your business but your endorsement of our products!

² Keep suspension on ice while in use.

³ See Appendix C for recommended seeding volumes.

Appendix A: Establishment of ObaCell® Cultures for In Vivo Implantation Workflow







Page 4 of 6 Obatala Sciences, Inc.

Appendix B: Troubleshooting

Problem	Reason	Solution	
No gelation observed	ObaGel [®] pre-warmed at	Do not heat ObaGel [®] prior	
	37°C or equivalent prior to	to use, maintain on ice	
	use	during use.	
	Repeated freeze-thaws	Aliquot ObaGel® into	
		smaller volumes and thaw	
		as needed.	
	Improper extended storage	Store at -20°C for up to	
		They and stars at 4°C for	
		naw and store at 4 C for	
Gelation during pipetting or	ObaGel [®] pre-warmed at	Do not heat ObaGel [®] prior	
handling	37°C or equivalent prior to	to use maintain on ice	
handing	use	during use.	
	ObaGel [®] not maintained at	Ensure ObaGel [®] is kept at	
	4°C or on ice prior to use	4°C or on wet ice prior to	
	•	use.	
Contraction of 3D	Physical disruption of the	Careful handling of the	
constructs	gel	plates and manipulation of	
		the constructs. Slowly	
		remove and add media	
		from edge of the well, tilt	
		the plate to reduce shear	
		stress at the gel-liquid	
		Interface.	
	Innomogeneous seeding	Pipet ObaGel®/ObaVate	
	solution	immediately prior to	
		solution	
		appears homogenous	
		Pinette solution while	
		seeding, every 3-6	
		replicates.	
Rapid spheroid or organoid	Too much media	Reduce media volume.	
formation in 3D constructs	Initial seeding density of	Reduce seeding density.	
	cells is too high	<u> </u>	
	Inhomogeneous seeding	Pipet several times to mix	
	solution	well and pipette several	
		times between replicates.	

Appendix C: Recommended Volumes

Reagent	6-well (5 mL)	24-well (1 mL)	96-well (100 μL)
ObaGel [®]	1.25 mL/well	250 µL/well	25 µL/well
ObaVate™	3.75 mL/well	750 µL/well	75 µL/well
Total Construct Volume	5 mL/well	1 mL/well	100 µL/well
hSVF cells	5,000,000 cells/well	1,000,000 cells/well	100,000 cells/well
hASC cells	500,000 cells/well	50,000-100,000 cells/well	5,000-10,000 cells/well
3D Medium Feedings	2.5 mL/well	500 μL/well	50 µL/well