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Obatala Sciences' Protocol 102 How Do I Harvest Adherent Cells from Obatala Sciences?

Written by: Obatala Sciences' Scientific Team Last Updated: January 2024

Reagents, Materials, and Equipment

- Obatala Sciences' Human Adipose-Derived Stromal/Stem Cells (Catalog #OS-101) or equivalent cryopreserved primary cell product
- Obatala Sciences' StromaQual[™] Stromal Medium (Catalog #OS-001) or medium of choice
- Phosphate buffered saline (1X) or equivalent product
- 0.05% trypsin/EDTA Solution
- ♦ 70% ethanol
- Sterile paper towel or kimwipe
- Conical centrifuge tube
- Flask, multi-well plate, or equivalent plasticware suitable for cell culture
- BSL2 Biological Safety Cabinet
- Benchtop centrifuge
- Water bath or equivalent equipment
- 37°C, 5% CO₂ incubator
- Micropipette and tips
- Serological pipettes
- Pipette controller

General Requirements

- 1. All personnel should be trained and certified by the Principal Investigator regarding Universal Precautions and Handling of Bloodborne Pathogens.
- 2. All procedures should be conducted by investigators using appropriate personal protective equipment at all times. Any waste materials should be decontaminated (bleached) and disposed of using appropriate biohazard waste containers.
- 3. Wear protective eyewear during handling of cryovial(s).

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Protocol

Initial Handling of Obatala Sciences' Products

- 1. Purchase and receive Obatala Sciences' Human Adipose-Derived Stromal/Stem Cells (Catalog #OS-101) or equivalent cryopreserved primary cell product.
- 2. When you receive the package containing your Obatala Sciences' cellular product(s), remove the cryovial(s) of cells from the dry ice using appropriate safety procedures.
- 3. For immediate use, thaw and seed the cryovial of cells as described in Obatala Sciences' Protocol 101.
 - a. For intermediate storage, transfer the cryovial(s) into an appropriate freezing container for controlled cooling and place in a -80°C freezer.
 - b. For long term storage, transfer the cryovial(s) into a liquid nitrogen dewar.
- 4. Proceed with harvesting protocol as outlined below after seeding the primary cells at a recommended density of 10² to 3 x 10⁴ per square centimeter and feeding the cells with Obatala Sciences' StromaQual[™] Stromal Medium (Catalog #OS-001) or medium of choice until desired level of confluency is reached.

Additional Recommendations for Handling and Use of Obatala Sciences' Products

- 1. Obatala Sciences does not recommend passaging primary cell products beyond passage 3 (P3).
- 2. Obatala Sciences does not recommend exceeding 80% confluency between passages.

Harvesting Adherent Cells from Obatala Sciences' Primary Cell Products

- 1. Allow the cells to reach the desired level of confluence based on microscopic examination, where the optimal level of confluency should be empirically determined.
- 2. Transfer the flask(s) or plasticware of choice to a BSL2 biological safety cabinet.
- 3. Aspirate the culture media from the flask(s) or plasticware of choice.
- 4. Rinse the flask(s) or plasticware of choice twice with pre-warmed (37°C) phosphate buffered saline or equivalent product.
- 5. Add 0.05% trypsin/EDTA to the flask or plate. We recommend using 1 mL for roughly 40 to 50 square centimeters of surface area. Tilt the flask(s) or plasticware of choice to ensure that 0.05% trypsin/EDTA solution coats the entire growth area.
- 6. Incubate the flask(s) or plasticware of choice in a 37°C incubator with 5% CO₂ and 85% relative humidity for 5 minutes. After 5 minutes, observe the cells under a microscope to verify that cells have begun lifting from the surface and are floating in solution when tapped. If fewer than ~10% of cells remain attached, proceed with the next steps.
 - a. If >50% of cells appear still attached to the plate with little change in morphology or lifting when tapped, return to incubator for additional trypsinization. Observe every minute for signs of lifting until fewer than ~10% of cells remain attached.
 - b. Do not allow cells to incubate in 0.05% trypsin/EDTA solution for longer than 8-10 minutes regardless of adherence. Cell viability will be impacted with prolonged exposure to 0.05% trypsin/EDTA solution.

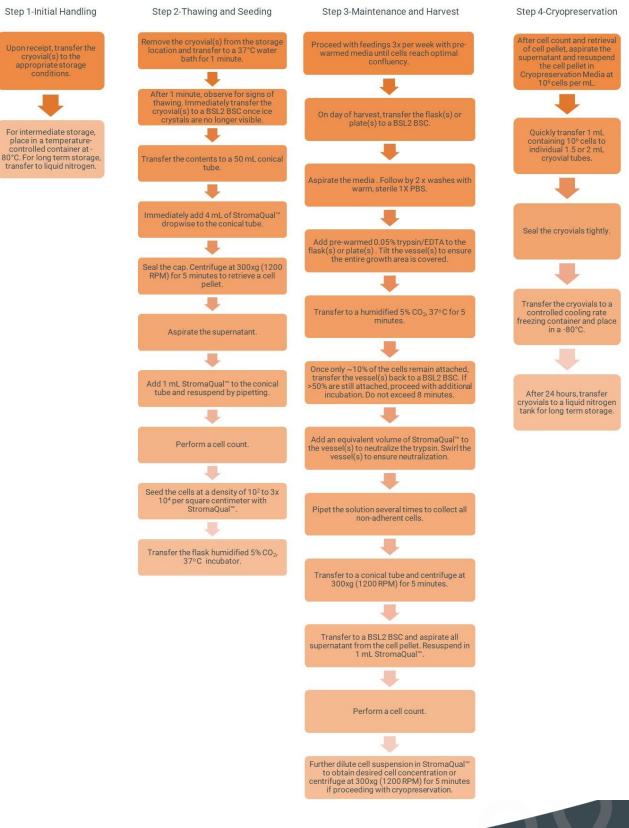
- 7. Return the flask(s) or plasticware of choice to the BSL2 biological safety cabinet.
- Add an equivalent volume of Obatala Sciences' StromaQual[™] Stromal Medium (Catalog #OS-001) or medium of choice to neutralize the volume of 0.05% trypsin/EDTA to each flask or plasticware of choice. Swirl flasks to ensure that solution is neutralized.
- 9. Pipet the solution inside the flask multiple times aseptically to collect all deadherent cells from the growth area.
- 10. Collect volume in a conical centrifuge tube.
- 11. Centrifuge the cells at 300x g (1200 rpm) at room temperature for 5 minutes.
- 12. Return conical centrifuge tube to the BSL2 Biological Safety Cabinet.
- 13. Carefully aspirate the supernatant from the cell pellet.
- 14. Resuspend pellet of cells in a volume of 1 mL of Obatala Sciences StromaQual[™] Stromal Medium (Catalog #OS-001) or medium of choice.
- 15. According to your laboratory's standard operating procedures, determine the relative percentage of live cells and dead cells to determine total live cells and viability (%).
 - a. A hemocytometer or automatic cell counter may be used.
 - b. For hSVF cells, we recommend an acridine orange/propidium iodide (AO/PI) viability stain.
 - c. For hASCs, we recommend trypan blue viability stain.
- 16. Further dilute the cell suspension in Obatala Sciences' StromaQual[™] Stromal Medium (Catalog #OS-001) or medium of choice at the desired cell concentration for culture expansion or centrifuge the cell suspension at 300x g (1200 rpm) for 5 minutes at room temperature to retrieve a cell pellet in preparation for cryopreservation or other procedures.

Recommended Protocols

Obatala Sciences' Protocol 101-How Do I Thaw Cryovials from Obatala Sciences? Obatala Sciences' Protocol 103-How Do I Cryopreserve Culture-Expanded Cells from Obatala Sciences?

Remember, any laboratory that mentions Obatala Sciences' products by name in a publication is eligible for a 10% discount on their next order! We appreciate not only your business but your endorsement of our products!

Appendix A: Handling, Thawing, Seeding, Harvesting, and Cryopreservation of Obatala Sciences' Cells Workflow



Appendix B: Troubleshooting

| Problem | Reason | Solution |
|---|---|--|
| Cells are adherent after 5 minutes incubation with 0.05% Trypsin/EDTA | Cells over overconfluent | Extend the incubation period to 7 minutes (do not exceed 8-10 minutes) |
| | Cells were not washed with 1X PBS prior to trypsinization | Wash the flask with warm, sterile 1X PBS prior to trypsinization |
| | Trypsin has undergone multiple heating cycles | Use a new lot of trypsin and/or store smaller aliquots to avoid multiple heating cycles |